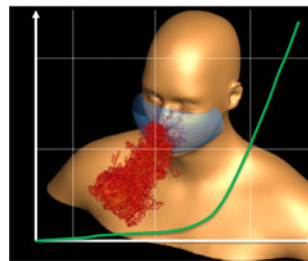


The flow physics of COVID-19

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The graphical abstract displays the growth of COVID-19 infections worldwide as of April 20, 2020, superimposed on the result from a direct-numerical simulation by Jung-Hee Seo (Johns Hopkins University) and Kourosh Shoele (Florida State University), showing the vortices generated by a cough through a face mask.

Flow physics plays a key role in nearly every facet of the COVID-19 pandemic. This includes the generation and aerosolization of virus-laden respiratory droplets from a host, its airborne dispersion and deposition on surfaces, as well as the subsequent inhalation of these bioaerosols by unsuspecting recipients. Fluid dynamics is also key to preventative measures such as the use of face masks, hand washing, ventilation of indoor environments and even social distancing. This article summarizes what we know and, more importantly, what we need to learn about the science underlying these issues so that we are better prepared to tackle the next outbreak of COVID-19 or a similar disease.

Key words: biomedical flows, aerosols/atomization, particle/fluid flow

1. Introduction

Transmission of respiratory infections such as COVID-19 is primarily via virus-laden fluid particles (i.e. droplets and aerosols) that are formed in the respiratory tract of an infected person and expelled from the mouth and nose during breathing, talking, coughing and sneezing (Jones & Brosseau 2015; Asadi *et al.* 2020; Bourouiba 2020; CDC 2020a). Wells (1934, 1955) showed that the competing effects of inertia, gravity and evaporation determine the fate of these droplets. Droplets larger than a critical size settle faster than they evaporate, and so contaminate surrounding surfaces. Droplets smaller than this size evaporate faster than they settle, so forming droplet nuclei that can stay airborne for hours and may be transported over long distances.

Human-to-human transmission of COVID-19 occurs primarily via three routes: large droplets that are expelled with sufficient momentum so as to directly impact the recipients' mouth, nose or conjunctiva; physical contact with droplets deposited on a surface and subsequent transfer to the recipient's respiratory mucosa; and inhalation

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by the recipient of aerosolized droplet nuclei from the expiratory ejecta that are delivered by ambient air currents. The first two routes associated with large droplets are referred to as the ‘droplet’ and ‘contact’ routes of transmission, whereas the third is the so-called ‘airborne’ transmission route (Jones & Brosseau 2015). Respiratory infections hijack our respiratory apparatus to increase the frequency and intensity of expiratory events, such as coughing and sneezing, which are particularly effective in generating and dispersing virus-carrying droplets.

Each stage in the transmission process is mediated by complex flow phenomena, ranging from air–mucous interaction, liquid sheet fragmentation, turbulent jets, and droplet evaporation and deposition, to flow-induced particle dispersion and sedimentation. Thus, flow physics is central to the transmission of COVID-19. Furthermore, given the importance of flow phenomena to the transmission process, the methods, devices and practices employed to mitigate respiratory infections are also rooted in the principles of fluid dynamics. These include simple methods such as hand washing and wearing face masks, to fogging machines, ventilation (Tang *et al.* 2006) and even practices such as social distancing. However, despite the long history of medical research and experience in the transmission of respiratory infections (a fascinating account of the ‘Spanish flu’ can be found in Soper (1919)), the rapid advance of COVID-19 around the world has laid bare the limits of our knowledge regarding the physics underlying the transmission process, as well as the inadequacy of the methods, devices and practices used to curtail transmission rates. For instance, one factor that is contributing to the rapid growth of COVID-19 infections is the higher viral load of the SARS-CoV-2 virus in the upper respiratory tract of asymptomatic hosts who shed virus-laden droplets during normal activities such as talking and breathing (Bai *et al.* 2020). This knowledge gap has also manifested through guidelines on practices such as social distancing and the wearing of face masks (Dwyer & Aubrey 2020; Elegant 2020), which are based on outdated science (Asadi *et al.* 2020; Bourouiba 2020).

This article attempts to summarize our current state of knowledge regarding the flow physics implicated in the transmission of COVID-19. The challenge of summarizing such a vast topic is amplified by the need of the hour, that speed take precedence over comprehensiveness. Readers are therefore referred to other articles on this topic (Tang *et al.* 2006; Xie *et al.* 2007; Johnson & Morawska 2009; Tang *et al.* 2009; Jones & Brosseau 2015; Asadi *et al.* 2020; Bourouiba 2020) to fill the many gaps that are sure to be left by this article.

2. Respiratory droplets and aerosols

This section addresses the generation, expulsion, evolution and transport of droplets and aerosols generated from the respiratory tract during expiratory activities such as breathing, talking, coughing and sneezing. The primary objective of fluid dynamic analyses in this setting is to: (a) determine the mechanisms for the generation of these droplets within the respiratory tract; (b) characterize the number density, size distribution and velocity of ejected droplets; (c) determine the critical droplet size for transition between the large and small droplet transmission routes; (d) estimate the settling distance of large droplets; (e) determine the evaporation times of small droplets; (f) characterize the transport of small droplets and droplet nuclei in the air; and (g) quantify the effect of external factors such as air currents, temperature and humidity on all of the above.

2.1. Mechanisms of droplet formation

It is generally established that respiratory droplets are formed from the fluid lining of the respiratory tract (Edwards *et al.* 2004; Morawska *et al.* 2005, 2009; Johnson & Morawska 2009; Almstrand *et al.* 2010). The mechanisms of formation are usually associated with distinct locations in the respiratory tract; this is important because both the characteristics of the respiratory tract (length scales, airway elasticity, mucus and saliva properties, etc.) as well as the viral load carried by the lining are functions of the location (Almstrand *et al.* 2010; Johnson *et al.* 2011).

One key mechanism for the generation of respiratory droplets is the instability (Moriarty & Grotberg 1999) and eventual fragmentation of the mucus lining due to the shear stress induced by the airflow. Predicting the fragmentation and droplet size distribution resulting from this fragmentation is non-trivial because mucus is a viscoelastic shear-thinning fluid subject to surface tension. This enables multiple instabilities to bear on this problem (Malashenko, Tsuda & Haber 2009), including surface-tension-driven Rayleigh–Plateau instability (Eggers 1997; Lin & Reitz 1998; Romanò *et al.* 2019), shear-driven Kelvin–Helmholtz instability (Kataoka, Ishii & Mishima 1983; Scardovelli & Zaleski 1999) and acceleration-driven Rayleigh–Taylor instability (Joseph, Beavers & Funada 2002; Halpern & Grotberg 2003). The Rayleigh–Taylor instability is particularly important in spasmodic events such as coughing and sneezing.

The second mechanism for droplet formation is associated with the rupture of the fluid lining during the opening of a closed respiratory passage (Malashenko *et al.* 2009). One important site for this mechanism is in the terminal bronchioles during breathing. These submillimetre-sized passages collapse during exhalation, and the subsequent reopening during inhalation ruptures the mucus meniscus, resulting in the generation of micrometre-sized droplets (Almstrand *et al.* 2010; Johnson *et al.* 2011). A similar mechanism probably occurs in the larynx during activities such as talking and coughing, which involve the opening and closing of the vocal folds (Mittal, Erath & Plesniak 2013). Finally, movement and contact of the tongue and lips, particularly during violent events such as sneezing, generate salivary droplets via this mechanism. The fluid dynamics of meniscus breakup associated with this mechanism is difficult to predict, especially given the non-Newtonian properties of the fluids involved, the dominant role of moving boundaries, and the large range of length and time scales implicated in this phenomenon.

2.2. Droplet characteristics

The number density, velocity and size distributions of droplets ejected by expiratory events have important implications for transmission, and numerous studies have attempted to measure these characteristics (Duguid 1946; Wells 1955; Morawska *et al.* 2009; Xie *et al.* 2009; Han, Weng & Huang 2013; Bourouiba, Dehandschoewercker & Bush 2014; Scharfman *et al.* 2016; Asadi *et al.* 2019). A single sneeze can generate $O(10^4)$ or more droplets, with velocities upwards of 20 m s^{-1} (Han *et al.* 2013). Coughing generates 10–100 times fewer droplets than sneezing, with velocities of approximately 10 m s^{-1} , but even talking can generate approximately 50 particles per second (Asadi *et al.* 2019). Measured droplet sizes range over four orders of magnitude, from $O(0.1)$ to $O(1000) \mu\text{m}$. Recent studies have noted that, while breathing generates droplets at a much lower rate, it probably accounts for more expired bioaerosols over the course of a day than intermittent events such as coughing and sneezing (Fiegel, Clarke & Edwards 2006; Atkinson & Wein 2008).

Consensus on all these droplet characteristics continues to be elusive due to the multifactorial nature of the phenomena as well as the difficulty of making such measurements (Chao *et al.* 2009; Morawska *et al.* 2009; Han *et al.* 2013).

2.3. The expiratory jet and droplet transmission

Droplets generated within the respiratory tract by the mechanisms described above are carried outwards by the respiratory airflow, and those droplets that are not reabsorbed by the fluid lining are expelled within a two-phase buoyant jet from the mouth and/or nose. Breathing and talking generate jet velocities that seldom exceed 5 m s^{-1} (Tang *et al.* 2013) and mostly expel small droplets. Violent expiratory events like coughing and sneezing, on the other hand, generate turbulent jets with Reynolds numbers of $O(10^4)$ and higher (Bourouiba *et al.* 2014). Mucus and saliva that are expelled out of the nose and mouth can be stretched into ligaments and sheets, and eventually fragment into small droplets if the Weber number is large enough (Jain *et al.* 2015). This breakup process probably contributes to the generation of large droplets that fall ballistically and contaminate nearby surfaces (Bourouiba *et al.* 2014).

Wells' simple but elegant analysis predicted that the critical size that differentiates large from small droplets is approximately $100 \mu\text{m}$ (Wells 1934). Subsequent analysis has shown that typical temperature and humidity variations expand the critical size range from approximately 50 to $150 \mu\text{m}$ (Xie *et al.* 2007). For the droplet transmission route, an important consideration is the horizontal distance travelled by the large droplets. Thus the 3–6 feet social distancing guidelines (CDC 2020b; WHO 2020) probably originate from Wells' original work. However, studies indicate that, while this distance might be adequate for droplets expelled during breathing and coughing (Xie *et al.* 2007; Wei & Li 2015), large droplets expelled from sneezes may travel 20 feet or more (Xie *et al.* 2007; Bourouiba *et al.* 2014). Studies also suggest that social distancing in indoor environments (Wong *et al.* 2004) could be complicated by ventilation-system-induced air currents.

It has also been shown that the respiratory jet transforms into a turbulent cloud or puff (Bourouiba *et al.* 2014). While large droplets are mostly not affected by the cloud dynamics, small and medium-sized droplets can be suspended in the turbulent cloud for a longer time by its circulatory flow, thereby extending the travel distance significantly (Bourouiba *et al.* 2014). This also has important implications for transmission via indirect contact with contaminated surfaces, since SARS-CoV-2 is able to survive on many types of surfaces for hours (van Doremalen *et al.* 2020). The turbulent cloud also moves upwards due to buoyancy (Bourouiba *et al.* 2014), thereby enabling small droplets and droplet nuclei to reach heights where they can enter the ventilation system and accelerate airborne transmissions (see §2.5). The notion of a critical droplet size that was introduced by Wells (1934) might need to be re-examined in the light of our rapidly evolving knowledge about these expiratory events (Xie *et al.* 2007; Bourouiba *et al.* 2014).

2.4. Droplet evaporation and droplet nuclei

Droplet evaporation plays a singularly important role in the eventual fate of a droplet (Wells 1934). The rate of evaporation depends on the difference between the droplet surface saturation vapour pressure and the vapour pressure of the surrounding air, the latter being dependent on humidity. The evaporation rate also depends on the mass-diffusion coefficient, which is a strong function of surface-to-ambient temperature difference, as well as the relative velocity between the droplet and

surrounding gas. Thus, Reynolds, Nusselt and Sherwood numbers for the droplets are just some of the non-dimensional parameters that determine this phenomenon (Xie *et al.* 2007). This dependence of evaporation rates on the ambient temperature and humidity has implications for the very important, and as yet unresolved, questions regarding seasonal and geographic variations in transmission rates (Tang 2009; Ma *et al.* 2020) as well as transmission in various indoor environments (Tang *et al.* 2006; Li *et al.* 2007).

Higher temperatures and lower relative humidities lead to larger evaporation rates that increase the critical droplet size (Wells 1934; Xie *et al.* 2007). However, temperature changes are usually accompanied by changes in humidity, and the overall effect of environmental conditions on transmission rates has been difficult to ascertain. This is not only due to the fact that these factors modulate the relative importance of the droplet and airborne routes of transmission, but also because survivability of enveloped viruses such as SARS-CoV-2 seem to be linked to these factors in a complex, non-monotonic manner (Geller, Varbanov & Duval 2012). Models that can combine droplet/aerosol fluid dynamics with virus microbiology and/or population dynamics could help unravel this complex effect of ambient conditions on transmission rates.

2.5. Airborne transmission

The airborne transmission route is associated with small droplets that are suspended and transported in air currents. Most of these droplets evaporate within a few seconds (Xie *et al.* 2007) to form droplet nuclei, although the vapour-rich, buoyant turbulent expiratory jet can slow this evaporation process (Bourouiba 2020). The nuclei consist of virions and solid residue (Vejerano & Marr 2018), but water may never be completely removed (Mezhericher, Levy & Borde 2010). These droplet nuclei are submicrometre to approximately 10 μm in size, and may remain suspended in the air for hours. Each droplet nucleus could contain multiple virions, and, given the approximately one hour viability half-life of the SARS-CoV-2 virus (van Doremalen *et al.* 2020) and the fact that SARS-type infections in a host may potentially be caused by a single virus (Nicas, Nazaroff & Hubbard 2005), droplet nuclei play a singularly important role in the transmission of COVID-19-type infections (Asadi *et al.* 2020). The evaporation process of virus-laden respiratory droplets and the composition of droplet nuclei require further analysis because these have implications for the viability and potency of the virus that is transported by these nuclei.

The transport of droplet nuclei over larger distances is primarily driven by ambient flows, and indoor environments such as homes, offices, malls, aircraft and public transport vehicles pose a particular challenge for disease transmission. The importance of ventilation in controlling airborne transmission of infections is well known (Tang *et al.* 2006; Li *et al.* 2007) and much of the recent work in this area has exploited the power of computational fluid dynamic (CFD) modelling (Thatiparti, Ghia & Mead 2017; Yang *et al.* 2018; Yu, Mui & Wong 2018). However, indoor spaces can have extremely complex flows, due not only to the presence of recirculatory flows driven by ventilation systems but also to anthropogenic thermally driven flow effects (Craven & Settles 2006; Licina *et al.* 2014). COVID-19 transmission from asymptomatic hosts (Bai *et al.* 2020; Ye *et al.* 2020) makes it more critical than ever that we develop methods of analysis that provide better prediction of these effects.

3. Inhalation and deposition of bioaerosols

The process of inhalation of virus-laden particles/droplets and deposition of the virus in the respiratory mucosa of the host is the final stage of airborne transmission. Fortunately, particle transport and deposition in the human airway has been studied extensively in the context of drug delivery (Heyder 2004), food smell (Ni *et al.* 2015) and pollutant transport (Morawska *et al.* 2005). The deposition of a solid particle is governed primarily by the mechanism of transport, whereas for liquid aerosols the evaporation/diffusion process contributes significantly to the deposition mechanism. The latter is, however, a complex subject and has not been studied extensively so far (Rostami 2009). There are six mechanisms that determine the deposition location: impaction, sedimentation, interception, diffusion, electrostatic precipitation and convection (Hinds 1999). The relative importance of these mechanisms depends on the particle size and the region of the airway where deposition occurs. In general, for larger particles, impaction, sedimentation and interception are more important than diffusion and convection (Rostami 2009). For droplet-nuclei-sized particles, sedimentation will drive significant deposition in the upper respiratory tract of the host (Willeke, Baron & Martonen 1993).

Deposition of virus-bearing droplets in the respiratory tract does not always result in infection, since the mucus layer provides some level of protection against virus invasion and subsequent infection (Zanin *et al.* 2016). The rate of droplet/nuclei deposition in the respiratory tract is quantified by the non-dimensionalized deposition velocity (Friedlander & Johnstone 1957; Liu & Agarwal 1974), which can vary by over four orders of magnitude (Guha 2008). For small droplets, deposition relies completely on turbulent diffusion (Friedlander & Johnstone 1957), but for large droplets, the deposition velocity increases substantially due to impact on the highly curved and complex passage walls of the respiratory tract. Large droplets, despite a higher deposition velocity, probably deposit in the upper respiratory system, and could be deactivated by the first defensive layer of the mucosa (Fokkens & Scheeren 2000). On the other hand, droplet nuclei, despite their smaller deposition velocity, will penetrate deeper into the respiratory system, and this could affect the progression and intensity of the infection.

Imaging modalities such as computed tomography (CT) and magnetic resonance imaging (MRI) provide realistic anatomical models for experiments (Ni *et al.* 2015) and CFD models (Rostami 2009), from which local deposition can be quantified. A recent study even included the immune system response in the model (Haghnegahdar, Zhao & Feng 2019), and similar models that combine fluid dynamics, biomechanics and virology could serve as important tools in combating such pandemics.

4. Measures to mitigate transmission

4.1. Mucus property modification

The physical properties of the mucus play a key role in droplet formation within the respiratory tract. Transient modification of the physical properties of the mucus lining via material delivery to enhance mucus stability therefore provides a means for reducing infection rates. Fiegel *et al.* (2006) used isotonic saline to change the mucus lining properties via the induced ionic charge to reduce droplet formation, and Edwards *et al.* (2004) explored the use of surface-tension-enhancing inhalants to reduce droplet generation. These techniques involve complex multiphysics flow phenomena that could benefit from advanced experimental and computational techniques.

4.2. Fogging machines

Fogging machines provide an effective means for disinfecting large spaces, such as hospitals, nursing homes, grocery stores and airplanes. Fogging machines that rely on the dispersion of a fine mist of disinfectants in the air have proven their performance in the healthcare sector (Otter *et al.* 2013) and the food industry (Oh *et al.* 2005). Commercial fogging machines are also designed based on the same flow physics of aerosolization, and their droplet size is below 10 μm (Krishnan *et al.* 2012) in order to facilitate extended airborne duration. For this range of droplet sizes, it is likely that inertial effects are small, and the collision between these disinfectant aerosol droplets and virus-bearing droplet nuclei might be dominated by diffusion. It is unclear if the collision rate will be enhanced by the turbulence generated by heating, ventilation and air-conditioning systems. Given that fogging machines have been widely employed in particle image velocimetry (PIV), experiments would be particularly well poised to study these phenomena.

4.3. Hand washing

Transmission of infection from surfaces with virion-laden respiratory droplets usually occurs via hands (Nicas & Jones 2009), and hand washing with soap therefore remains the most effective strategy for mitigating this mode of transmission (Stock & Francis 1940). Soap molecules have a polar ionic hydrophilic side and a non-polar hydrophobic side that bonds with oils and lipids. Hand washing therefore works by emulsifying the lipid content of the material adhering to the hand into the bulk fluid and convecting it away. For enveloped viruses such as SARS-CoV-2, soap molecules also dismantle the lipid envelope of the virus, thereby deactivating it (Kohn, Gitelman & Inbar 1980). The detritus from this disintegration is then trapped by the soap molecules into micelles, which are washed away.

These molecular-level mechanisms are powered by macro-level flow phenomena associated with the movements of the hands. Amazingly, despite the 170+ year history of hand washing in medical hygiene (Rotter 1997), we were unable to find a single published research article on the flow physics of hand washing. The relative movement of the hands generates complex shear-driven flows of the soapy water, which forms a foam-laden, multiphase emulsion. Soap bubbles, which trap micelles, segregate rapidly from the fluid phase, thereby further accelerating the removal process.

It is known that liquid foam exhibits elastic and plastic deformation under small and large stresses, respectively. With large enough deformation rates, the foam can rearrange its network and flow (Weaire & Hutzler 1999), which can be studied experimentally (Janiaud & Graner 2005). Reynolds numbers for these soapy liquid layers could exceed $O(1000)$, suggesting that inertia, viscosity, surface tension and gravitational forces would all play an important role in this process. An improved understanding of hand-washing flow physics in the COVID-19 era could provide a science-based foundation for public guidelines/recommendations as well as new technologies that could improve the effectiveness of this practice.

4.4. Face masks

One issue that has generated significant controversy during the COVID-19 pandemic is the effectiveness of face masks (Dwyer & Aubrey 2020; Elegant 2020; Feng *et al.* 2020). Indeed, it is likely that the years ahead will see the use of face masks become

a norm in our lives. Understanding the physics that underpins the effectiveness of face masks as a defence against airborne pathogens is, therefore, more important than ever.

Face masks provide ‘inward’ protection by filtering virus-laden aerosolized particles that would otherwise be inhaled by an uninfected person, and ‘outward’ protection by trapping virus-laden droplets expelled by an infected person (van der Sande, Teunis & Sabel 2008). The effectiveness of a simple face mask such as the surgical, N95 or homemade cloth face mask is a function of the combined effect of the filtering properties of the face-mask material, the fit of the mask on the face and the related leaks from the perimeter of the face mask. Each of these features implicates complex flow phenomena, which are briefly addressed here.

4.4.1. Inward protection

The face-mask material traps droplets and particles via the combined effects of diffusion, inertial impaction, interception and electrostatic attraction (Thomas *et al.* 2016; Fleming 2020). Filter efficiency (the ratio of the particle concentrations upstream and downstream of the mask) is a function of the particle- and fibre-size-based Reynolds numbers, fibre-based Péclet number (for diffusion), particle-to-fibre size ratio (for interception) and Stokes number (for impaction). The nonlinear variation of filtration mechanisms on these parameters generates a complex dependence of the filter efficiency on flow velocity, particle size and filter material characteristics such as pore size, fibre diameter and electrostatic charge.

The process of inhalation generates a low pressure in the region interior to the face mask, thereby sealing (or at least reducing) perimeter leaks. Thus, with a reasonably well-fitted face mask, inward protection depends primarily on the face-mask filter material. In this regard, an important characteristic of a face mask is the dependence of filtration efficiency on particle size. Studies (Chen & Willeke 1992; Weber *et al.* 1993; Balazy *et al.* 2006) have shown that, for a given filter, there is an intermediate particle size where filtration efficiency is minimum. Below this size, electrostatic attraction (active for masks such as N95) and diffusion dominate filtration, whereas above this size, impaction and interception are the dominant mechanisms. Aerosolized virus-laden droplets and droplet nuclei vary in size from submicrometre to millimetres, and therefore the aforementioned size-dependent filtration efficiency is an important consideration for inward protection against COVID-19 infections. An increase in fibre density to enhance filtration efficiency is accompanied by an increased pressure drop across the mask (Lai, Poon & Cheung 2012), which requires a greater inhalation effort by the wearer. Thus, an appropriate balance has to be achieved via proper design of the filter material, and this might be particularly important in the post-COVID-19 era, if wearing face masks becomes routine.

4.4.2. Outward protection

The outward protection afforded by face masks has emerged as a particularly important issue in the COVID-19 pandemic because a SARS-CoV-2 transmission may occur early in the course of infection, not only from symptomatic patients but also from asymptomatic as well as minimally symptomatic patients (Bai *et al.* 2020; Ye *et al.* 2020; Zou *et al.* 2020). Indeed, the late switch to recommending universal use of face masks in the USA (Dwyer & Aubrey 2020; Elegant 2020) was based on the recognition that this spread by asymptomatic hosts might be a significant driver of COVID-19 infections.

While a mask can significantly reduce the velocity of the throughflow jet during expiratory events (Tang *et al.* 2009), the increased pressure in the region between the

mask and the face pushes the face mask outwards, resulting in increased perimeter leakage (Liu *et al.* 1993; Lei *et al.* 2013). This fluid–structure interaction problem is mediated by the structural design as well as the permeability of the mask. The leakage jets that issue from the perimeter can be turbulent and highly directed (see, for example, the flow visualization in Tang *et al.* (2009)), potentially serving as effective dispersers of respiratory aerosols in transverse directions. Spasmodic expiratory events such as coughing and sneezing that generate high transient expulsion velocities will significantly diminish the outward protection effectiveness of face masks (Lai *et al.* 2012). However, in a conceivable future where people will wear face masks while engaged in their daily routines, mask effectiveness during normal activities such as breathing and talking might be equally important.

In contrast to the problem of inward protection, which has been studied extensively (Chen & Willeke 1992; Weber *et al.* 1993; Bałazy *et al.* 2006), the flow physics of outward protection from face masks is less well studied. Tang *et al.* (2009) used Schlieren imaging to visualize cough-induced flow with and without face masks (surgical and N95). The study was extremely inventive but mostly qualitative, and future experiments should provide quantitative analysis of the leakage and throughflow jets, the aerosol dispersion through these jets, as well as the deformation of the mask during a variety of expiratory events. Recent CFD studies of face-mask aerodynamics (Lei *et al.* 2013; Zhu *et al.* 2016) demonstrate the potential of computational modelling for this problem, but there is a critical need for modelling flow-induced billowing and associated leakage enhancement during expiratory events. Ultimately, analysis should not only enable a detailed evaluation of the protective efficiency of face masks; it should also drive design changes that enhance mask performance and provide data that inform guidelines on practices such as social distancing.

5. Closing

The COVID-19 pandemic has exposed significant scientific gaps in our understanding of critical issues, ranging from the transmission pathways of such respiratory diseases, to the strategies to use for mitigating these transmissions. This article summarizes a fluid dynamicist's perspective on important aspects of the problem, including respiratory droplet formation, two-phase expiratory flows, droplet evaporation and transport, and face-mask aerodynamics. COVID-19 touches almost every major arena of fluid dynamics, from hydrodynamic instability to porous-media and turbulent shear flows, from droplet breakup to particle deposition, and from Newtonian gas flows to non-Newtonian liquids. For the topics that we have discussed, breadth and epidemiological context have taken precedence over a detailed exposition. COVID-19 has thrust the field of fluid dynamics into the public eye in a way (Bourouiba 2020; Parshina-Kottas *et al.* 2020) not seen since the space race of the 1960s. Our hope is that not only will this article serve as a call-to-arms to fluid dynamicists, it will also provide a starting point for the researcher who is motivated to tackle the science of COVID-19, and other similar diseases that are sure to appear in the not-too-distant future.

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Declaration of interests

The authors report no conflict of interests.

References

- ALMSTRAND, A.-C., BAKE, B., LJUNGSTRÖM, E., LARSSON, P., BREDBERG, A., MIRGORODSKAYA, E. & OLIN, A.-C. 2010 Effect of airway opening on production of exhaled particles. *J. Appl. Physiol.* **108** (3), 584–588.
- ASADI, S., BOUVIER, N., WEXLER, A. S. & RISTENPART, W. D. 2020 The coronavirus pandemic and aerosols: does COVID-19 transmit via expiratory particles? *Aerosol Sci. Technol.* **54** (6), 635–638.
- ASADI, S., WEXLER, A. S., CAPPA, C. D., BARREDA, S., BOUVIER, N. M. & RISTENPART, W. D. 2019 Aerosol emission and superemission during human speech increase with voice loudness. *Sci. Rep.* **9** (1), 2348.
- ATKINSON, M. P. & WEIN, L. M. 2008 Quantifying the routes of transmission for pandemic influenza. *Bull. Math. Biol.* **70** (3), 820–867.
- BALAŻY, A., TOIVOLA, M., ADHIKARI, A., SIVASUBRAMANI, S. K., REPONEN, T. & GRINSHPUN, S. A. 2006 Do N95 respirators provide 95% protection level against airborne viruses, and how adequate are surgical masks? *Am. J. Infect. Control* **34** (2), 51–57.
- BAI, Y., YAO, L., WEI, T., TIAN, F., JIN, D.-Y., CHEN, L. & WANG, M. 2020 Presumed asymptomatic carrier transmission of COVID-19. *J. Am. Med. Assoc.* **323** (14), 1406–1407.
- BOUROUBA, L. 2020 Turbulent gas clouds and respiratory pathogen emissions: potential implications for reducing transmission of COVID-19. *J. Am. Med. Assoc.*, doi:10.1001/jama.2020.4756.
- BOUROUBA, L., DEHANDSCHOEWERCKER, E. & BUSH, J. W. M. 2014 Violent expiratory events: on coughing and sneezing. *J. Fluid Mech.* **745**, 537–563.
- CENTERS FOR DISEASE CONTROL (CDC) 2020a How COVID-19 Spreads. Available at: <https://www.cdc.gov/coronavirus/2019-ncov/prepare/transmission.html>.
- CENTERS FOR DISEASE CONTROL (CDC) 2020b CDC Guidelines on Social Distancing. Available at: <https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/social-distancing.html>.
- CHAO, C. Y. H., WAN, M. P., MORAWSKA, L., JOHNSON, G. R., RISTOVSKI, Z. D., HARGREAVES, M., MENSERSEN, K., CORBETT, S., LI, Y., XIE, X. & KATOSHEVSKI, D. 2009 Characterization of expiration air jets and droplet size distributions immediately at the mouth opening. *J. Aerosol Sci.* **40** (2), 122–133.
- CHEN, C.-C. & WILLEKE, K. 1992 Aerosol penetration through surgical masks. *Am. J. Infect. Control* **20** (4), 177–184.
- CRAVEN, B. A. & SETTLES, G. S. 2006 A computational and experimental investigation of the human thermal plume. *Trans. ASME J. Fluids Engng* **128**, 1251–1258.
- VAN DOREMALEN, N., BUSHMAKER, T., MORRIS, D. H., HOLBROOK, M. G., GAMBLE, A., WILLIAMSON, B. N., TAMIN, A., HARCOURT, J. L., THORNBURG, N. J., GERBER, S. I. *et al.* 2020 Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1. *New Engl. J. Med.* **382**, 1564–1567.
- DUGUID, J. P. 1946 The size and the duration of air-carriage of respiratory droplets and droplet-nuclei. *Epidemiol. Infect.* **44** (6), 471–479.
- DWYER, C. & AUBREY, A. 2020 CDC now recommends Americans consider wearing cloth face coverings in public. Available at: <https://www.npr.org/sections/coronavirus-live-updates/2020/04/03/826219824/president-trump-says-cdc-now-recommends-americans-wear-cloth-masks-in-public>.
- EDWARDS, D. A., MAN, J. C., BRAND, P., KATSTRA, J. P., SOMMERER, K., STONE, H. A., NARDELL, E. & SCHEUCH, G. 2004 Inhaling to mitigate exhaled bioaerosols. *Proc. Natl Acad. Sci. USA* **101** (50), 17383–17388.
- EGGERS, J. 1997 Nonlinear dynamics and breakup of free-surface flows. *Rev. Mod. Phys.* **69** (3), 865.

- ELEGANT, N. X. 2020 Coronavirus outbreak changes US's mind on everyone wearing face masks. *Fortune*. Available at: <https://fortune.com/2020/04/03/coronavirus-face-mask-cdc/>.
- FENG, S., SHEN, C., XIA, N., SONG, W., FAN, M. & COWLING, B. J. 2020 Rational use of face masks in the covid-19 pandemic. *Lancet Respir. Med.* Available at: [https://www.thelancet.com/journals/lanres/article/PIIS2213-2600\(20\)30134-X/fulltext](https://www.thelancet.com/journals/lanres/article/PIIS2213-2600(20)30134-X/fulltext).
- FIEGEL, J., CLARKE, R. & EDWARDS, D. A. 2006 Airborne infectious disease and the suppression of pulmonary bioaerosols. *Drug Discov. Today* **11** (1–2), 51–57.
- FLEMING, A. 2020 Keep it clean: the surprising 130-year history of handwashing. *The Guardian*. Available at: <https://www.theguardian.com/world/2020/mar/18/keep-it-clean-the-surprising-130-year-history-of-handwashing>.
- FOKKENS, W. J. & SCHEEREN, R. A. 2000 Upper airway defence mechanisms. *Paediat. Respir. Rev.* **1** (4), 336–341.
- FRIEDLANDER, S. K. & JOHNSTONE, H. F. 1957 Deposition of suspended particles from turbulent gas streams. *Ind. Engng Chem.* **49** (7), 1151–1156.
- GELLER, C., VARBANOV, M. & DUVAL, R. 2012 Human coronaviruses: insights into environmental resistance and its influence on the development of new antiseptic strategies. *Viruses* **4** (11), 3044–3068.
- GUHA, A. 2008 Transport and deposition of particles in turbulent and laminar flow. *Annu. Rev. Fluid Mech.* **40**, 311–341.
- HAGHNEGHAHDAR, A., ZHAO, J. & FENG, Y. 2019 Lung aerosol dynamics of airborne influenza A virus-laden droplets and the resultant immune system responses: an in silico study. *J. Aerosol Sci.* **134**, 34–55.
- HALPERN, D. & GROTTBERG, J. B. 2003 Nonlinear saturation of the Rayleigh instability due to oscillatory flow in a liquid-lined tube. *J. Fluid Mech.* **492**, 251–270.
- HAN, Z. Y., WENG, W. G. & HUANG, Q. Y. 2013 Characterizations of particle size distribution of the droplets exhaled by sneeze. *J. R. Soc. Interface* **10** (88), 20130560.
- HEYDER, J. 2004 Deposition of inhaled particles in the human respiratory tract and consequences for regional targeting in respiratory drug delivery. *Proc. Am. Thorac. Soc.* **1** (4), 315–320.
- HINDS, W. C. 1999 *Aerosol Technology: Properties, Behavior, and Measurement of Airborne Particles*. John Wiley and Sons.
- JAIN, M., PRAKASH, R. S., TOMAR, G. & RAVIKRISHNA, R. V. 2015 Secondary breakup of a drop at moderate Weber numbers. *Proc. R. Soc. Lond. A* **471** (2177), 20140930.
- JANIAUD, E. & GRANER, F. 2005 Foam in a two-dimensional Couette shear: a local measurement of bubble deformation. *J. Fluid Mech.* **532**, 243–267.
- JOHNSON, G. R. & MORAWSKA, L. 2009 The mechanism of breath aerosol formation. *J. Aerosol Med. Pulm. Drug Deliv.* **22** (3), 229–237.
- JOHNSON, G. R., MORAWSKA, L., RISTOVSKI, Z. D., HARGREAVES, M., MENSERSEN, K., CHAO, C. Y. H., WAN, M. P., LI, Y., XIE, X., KATOSHEVSKI, D. *et al.* 2011 Modality of human expired aerosol size distributions. *J. Aerosol Sci.* **42** (12), 839–851.
- JONES, R. M. & BROSSEAU, L. M. 2015 Aerosol transmission of infectious disease. *J. Occup. Environ. Med.* **57** (5), 501–508.
- JOSEPH, D. D., BEAVERS, G. S. & FUNADA, T. 2002 Rayleigh–Taylor instability of viscoelastic drops at high Weber numbers. *J. Fluid Mech.* **453**, 109–132.
- KATAOKA, I., ISHII, M. & MISHIMA, K. 1983 Generation and size distribution of droplet in annular two-phase flow. *Trans. ASME J. Fluids Engng* **105**, 230–238.
- KOHN, A., GITELMAN, J. & INBAR, M. 1980 Unsaturated free fatty acids inactivate animal enveloped viruses. *Arch. Virol.* **66** (4), 301–307.
- KRISHNAN, J., FEY, G., STANSFIELD, C., LANDRY, L., NGUY, H., KLASSEN, S. & ROBERTSON, C. 2012 Evaluation of a dry fogging system for laboratory decontamination. *Appl. Biosafety* **17** (3), 132–141.
- LAI, A. C. K., POON, C. K. M. & CHEUNG, A. C. T. 2012 Effectiveness of facemasks to reduce exposure hazards for airborne infections among general populations. *J. R. Soc. Interface* **9** (70), 938–948.

- LEI, Z., YANG, J., ZHUANG, Z. & ROBERGE, R. 2013 Simulation and evaluation of respirator face seal leaks using computational fluid dynamics and infrared imaging. *Ann. Occup. Hyg.* **57** (4), 493–506.
- LI, Y., LEUNG, G. M., TANG, J. W., YANG, X., CHAO, C. Y., LIN, J. Z., LU, J. W., NIELSEN, P. V., NIU, J., QIAN, H. *et al.* 2007 Role of ventilation in airborne transmission of infectious agents in the built environment – a multidisciplinary systematic review. *Indoor Air* **17** (1), 2–18.
- LICINA, D., PANTELIC, J., MELIKOV, A., SEKHAR, C. & THAM, K. W. 2014 Experimental investigation of the human convective boundary layer in a quiescent indoor environment. *Build. Environ.* **75**, 79–91.
- LIN, S. P. & REITZ, R. D. 1998 Drop and spray formation from a liquid jet. *Annu. Rev. Fluid Mech.* **30** (1), 85–105.
- LIU, B. Y. H. & AGARWAL, J. K. 1974 Experimental observation of aerosol deposition in turbulent flow. *J. Aerosol Sci.* **5** (2), 145–155.
- LIU, B. Y. H., LEE, J.-K., MULLINS, H. & DANISCH, S. G. 1993 Respirator leak detection by ultrafine aerosols: a predictive model and experimental study. *Aerosol Sci. Technol.* **19** (1), 15–26.
- MA, Y., ZHAO, Y., LIU, J., HE, X., WANG, B., FU, S., YAN, J., NIU, J., ZHOU, J. & LUO, B. 2020 Effects of temperature variation and humidity on the death of Covid-19 in Wuhan, China. *Sci. Total Environ.* **724**, 138226–138226.
- MALASHENKO, A., TSUDA, A. & HABER, S. 2009 Propagation and breakup of liquid menisci and aerosol generation in small airways. *J. Aerosol Med. Pulm. Drug Deliv.* **22** (4), 341–353.
- MEZHERICHER, M., LEVY, A. & BORDE, I. 2010 Theoretical models of single droplet drying kinetics: a review. *Dry. Technol.* **28** (2), 278–293.
- MITTAL, R., ERATH, B. D. & PLESNIAK, M. W. 2013 Fluid dynamics of human phonation and speech. *Annu. Rev. Fluid Mech.* **45**, 437–467.
- MORAWSKA, L., HOFMANN, W., HITCHINS-LOVEDAY, J., SWANSON, C. & MENGERSEN, K. 2005 Experimental study of the deposition of combustion aerosols in the human respiratory tract. *J. Aerosol Sci.* **36** (8), 939–957.
- MORAWSKA, L., JOHNSON, G. R., RISTOVSKI, Z. D., HARGREAVES, M., MENGERSEN, K., CORBETT, S., CHAO, C. Y. H., LI, Y. & KATOSHEVSKI, D. 2009 Size distribution and sites of origin of droplets expelled from the human respiratory tract during expiratory activities. *J. Aerosol Sci.* **40** (3), 256–269.
- MORIARTY, J. A. & GROTBORG, J. B. 1999 Flow-induced instabilities of a mucous bilayer. *J. Fluid Mech.* **397**, 1–22.
- NI, R., MICHALSKI, M. H., BROWN, E., DOAN, N., ZINTER, J., OUELLETTE, N. T. & SHEPHERD, G. M. 2015 Optimal directional volatile transport in retronasal olfaction. *Proc. Natl Acad. Sci. USA* **112** (47), 14700–14704.
- NICAS, M. & JONES, R. M. 2009 Relative contributions of four exposure pathways to influenza infection risk. *Risk Anal. Intl J.* **29** (9), 1292–1303.
- NICAS, M., NAZAROFF, W. W. & HUBBARD, A. 2005 Toward understanding the risk of secondary airborne infection: emission of respirable pathogens. *J. Occup. Environ. Hyg.* **2** (3), 143–154.
- OH, S.-W., GRAY, P. M., DOUGHERTY, R. H. & KANG, D.-H. 2005 Aerosolization as novel sanitizer delivery system to reduce food-borne pathogens. *Lett. Appl. Microbiol.* **41** (1), 56–60.
- OTTER, J. A., YEZLI, S., PERL, T. M., BARBUT, F. & FRENCH, G. L. 2013 The role of no-touch automated room disinfection systems in infection prevention and control. *J. Hosp. Infect.* **83** (1), 1–13.
- PARSHINA-KOTTAS, Y., SAGET, B., PATANJALI, K., FLEISHER, O. & GIANORDOLI, G. 2020 This 3-D simulation shows why social distancing is so important. *The New York Times*. Available at: <https://www.nytimes.com/interactive/2020/04/14/science/coronavirus-transmission-cough-6-feet-ar-ul.html>.
- ROMANÒ, F., FUJIOKA, H., MURADOGLU, M. & GROTBORG, J. B. 2019 Liquid plug formation in an airway closure model. *Phys. Rev. Fluids* **4** (9), 093103.
- ROSTAMI, A. A. 2009 Computational modeling of aerosol deposition in respiratory tract: a review. *Inhal. Toxicol.* **21** (4), 262–290.

- ROTTER, M. L. 1997 150 years of hand disinfection-Semmelweis' heritage. *Hyg. Med.* **22**, 332–339.
- VAN DER SANDE, M., TEUNIS, P. & SABEL, R. 2008 Professional and home-made face masks reduce exposure to respiratory infections among the general population. *PLoS ONE* **3** (7), e2618.
- SCARDOVELLI, R. & ZALESKI, S. 1999 Direct numerical simulation of free-surface and interfacial flow. *Annu. Rev. Fluid Mech.* **31** (1), 567–603.
- SCHARFMAN, B. E., TECHET, A. H., BUSH, J. W. M. & BOUROUIBA, L. 2016 Visualization of sneeze ejecta: steps of fluid fragmentation leading to respiratory droplets. *Exp. Fluids* **57** (2), 24.
- SOPER, G. 1919 The lessons of the pandemic. *Science* **49**, 501–506.
- STOCK, C. C. & FRANCIS, T. JR. 1940 The inactivation of the virus of epidemic influenza by soaps. *J. Exp. Med.* **71** (5), 661–681.
- TANG, J. W. 2009 The effect of environmental parameters on the survival of airborne infectious agents. *J. R. Soc. Interface* **6** (suppl_6), S737–S746.
- TANG, J. W., LI, Y., EAMES, I., CHAN, P. K. S. & RIDGWAY, G. L. 2006 Factors involved in the aerosol transmission of infection and control of ventilation in healthcare premises. *J. Hosp. Infect.* **64** (2), 100–114.
- TANG, J. W., LIEBNER, T. J., CRAVEN, B. A. & SETTLES, G. S. 2009 A schlieren optical study of the human cough with and without wearing masks for aerosol infection control. *J. R. Soc. Interface* **6** (suppl_6), S727–S736.
- TANG, J. W., NICOLLE, A. D., KLETTNER, C. A., PANTELIC, J., WANG, L., SUHAIMI, A. B., TAN, A. Y. L., ONG, G. W. X., SU, R., SEKCHAR, C. *et al.* 2013 Airflow dynamics of human jets: sneezing and breathing-potential sources of infectious aerosols. *PLoS ONE* **8** (4), e59970.
- THATIPARTI, D. S., GHIA, U. & MEAD, K. R. 2017 Computational fluid dynamics study on the influence of an alternate ventilation configuration on the possible flow path of infectious cough aerosols in a mock airborne infection isolation room. *Sci. Technol. Built Environ.* **23** (2), 355–366.
- THOMAS, D., CHARVET, A., BARDIN-MONNIER, N. & APPERT-COLLIN, J.-C. 2016 *Aerosol Filtration*. Elsevier.
- VEJERANO, E. P. & MARR, L. C. 2018 Physico-chemical characteristics of evaporating respiratory fluid droplets. *J. R. Soc. Interface* **15** (139), 20170939.
- WEAIRE, D. & HUTZLER, S. 1999 *The Physics of Foams*. Oxford University Press.
- WEBER, A., WILLEKE, K., MARCHLONI, R., MYOJO, T., MCKAY, R., DONNELLY, J. & LIEBHABER, F. 1993 Aerosol penetration and leakage characteristics of masks used in the health care industry. *Am. J. Infect. Control* **21** (4), 167–173.
- WEI, J. & LI, Y. 2015 Enhanced spread of expiratory droplets by turbulence in a cough jet. *Build. Environ.* **93**, 86–96.
- WELLS, W. F. 1934 On air-borne infections: study II. Droplets and droplet nuclei. *Am. J. Epidemiol.* **20** (3), 611–618.
- WELLS, W. F. 1955 Airborne contagion and air hygiene: an ecological study of droplet infections. *J. Am. Med. Assoc.* **159** (1), 90–90.
- WORLD HEALTH ORGANIZATION (WHO) 2020 Advice for public – maintain at least 1 metre (3 feet) distance between yourself and anyone who is coughing or sneezing. Available at: <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/advice-for-public>.
- WILLEKE, K., BARON, P. & MARTONEN, T. 1993 Aerosol measurement: principles, techniques and applications. *J. Aerosol. Med.* **6** (4), 317–320.
- WONG, T.-W., LEE, C.-K., TAM, W., LAU, J. T.-F., YU, T.-S., LUI, S.-F., CHAN, P. K. S., LI, Y., BRESEE, J. S., SUNG, J. J. Y. *et al.* 2004 Cluster of SARS among medical students exposed to single patient, Hong Kong. *Emerg. Infect. Dis.* **10** (2), 269–276.
- XIE, X., LI, Y., CHWANG, A. T. Y., HO, P. L. & SETO, W. H. 2007 How far droplets can move in indoor environments – revisiting the Wells evaporation-falling curve. *Indoor Air* **17** (3), 211–225.

- XIE, X., LI, Y., SUN, H. & LIU, L. 2009 Exhaled droplets due to talking and coughing. *J. R. Soc. Interface* **6** (suppl_6), S703–S714.
- YANG, L., LI, X., YAN, Y. & TU, J. 2018 Effects of cough-jet on airflow and contaminant transport in an airliner cabin section. *J. Comput. Multiphase Flows* **10** (2), 72–82.
- YE, F., XU, S., RONG, Z., XU, R., LIU, X., DENG, P., LIU, H. & XU, X. 2020 Delivery of infection from asymptomatic carriers of Covid-19 in a familial cluster. *Intl J. Infect. Dis.* **94**, 133–138.
- YU, H., MUI, K. & WONG, L. 2018 Numerical simulation of bioaerosol particle exposure assessment in office environment from MVAC systems. *J. Comput. Multiphase Flows* **10** (2), 59–71.
- ZANIN, M., BAVISKAR, P., WEBSTER, R. & WEBBY, R. 2016 The interaction between respiratory pathogens and mucus. *Cell Host Microbe* **19** (2), 159–168.
- ZHU, J. H., LEE, S. J., WANG, D. Y. & LEE, H. P. 2016 Evaluation of rebreathed air in human nasal cavity with N95 respirator: a CFD study. *Trauma Emerg. Care* **1** (2), 15–18.
- ZOU, L., RUAN, F., HUANG, M., LIANG, L., HUANG, H., HONG, Z., YU, J., KANG, M., SONG, Y., XIA, J. *et al.* 2020 SARS-CoV-2 viral load in upper respiratory specimens of infected patients. *New Engl. J. Med.* **382** (12), 1177–1179.